

**Master in Laboratory Animal Science and Welfare**

**Introduction**

The UAB Master in Laboratory Animal Science & Welfare has been running since 2000 to provide specialized training for professionals who work with animals for research purposes.

The program is accredited by the

[Federation of European Laboratory Animal Science Associations](http://www.felasa.eu/)



and forms part of the UAB Laboratory Animal Residential Program recognised by

[European College for Laboratory Animal Medicine](https://eclam.eu/)



For Veterinarians, completion of the 6 master modules allows to obtain the Veterinarian Certificate in Laboratory Animal Science and Medicine, approved by VetCEE [Veterinary Continuous Education in Europe (VetCEE)](https://www.vetcee.eu/)



**Course Structure:**

The course comprises 6 modules, a practical placement and a project. It is designed to be completed in three years and to be fully compatible with a full time job. The modules are intensive and two are run each year lasting between 1-2 weeks. In this way the course can accommodate students who live outside Barcelona.

Students are individually evaluated for each module.

 **Course Objectives:**

The program aims to provide expertise and professional excellence in all areas of laboratory animal science relevant to animal experimentation.

Specifically, the program is designed for veterinarians and other specialist personnel designated with responsibility for the welfare and care of laboratory animals.

Each module can be completed independently with some modules leading to accreditation for some of the functions described in EU Directive EU63/2010 and DR 53/2013 such as performing procedures, project design and project evaluation. See individual modules for details.

Completion of the whole course enables students to carry out functions described in EU Directive EU63/2010. For details, see:

European Commission, Directorate-General for Environment, *Caring for animals aiming for better science – Directive 2010/63/EU on protection of animals used for scientific purposes : education and training framework*, Publications Office, 2019, <https://data.europa.eu/doi/10.2779/311480>

**Registration**

Students may register for individual modules or for the whole Master course.

Special reduced fees are applicable for members of SECAL, ESLAV or ECLAM Residents.

In some cases, students may be eligible for a grant from Laboratory Animals or ICLAS.

Alternatively, students may register for individual modules each of which may lead to an official UAB certificate which can be used to demonstrate competence in the areas covered by the module.
**Course Director:**

 Professor PatriVergara (DipECLAM); Department of Cellular Biology, Physiology and Immunology. Faculty of Veterinary Science, Universitat Autònoma de Barcelona (tel: +34 935 811 848; e-mail: patri.vergara@uab.es)

**Organising Faculty/Department:**

 Department of Cellular Biology, Physiology and Immunology. Faculty of Veterinary Science, Universitat Autònoma de Barcelona.
More information: pg.animals.laboratori@uab.cat

**Modules detail:**

Each module consists of webinars and face to face lectures and practical sessions.

Following are the details of each module and estimated dates for the next edition of each module. Some topics and teachers may change as each module is updated for each edition. Details syllabus and specific dates are fixed at least 6 months in advance to the start of the module.

**Module:**

**Animal Experimentation: Welfare, Legislation, Ethical Committees and Evaluation of Projects**

The course focuses on the topics of Welfare, Ethical Committees and Legislation in respect of laboratory animals.

This module provides the knowledge and skills required by designated veterinarians and other onsite personnel responsible for the welfare and care of animals to perform the functions of project evaluation.

The content of this module also complies with the training requirements set out in European Directive EU63/2010, developed by the Expert Working Group, supported by European Commission, in respect of the following modules: 1) National Legislation; 2) Ethics, welfare and the 3Rs (level 1); 5) Recognition of pain, suffering and distress; 9) Ethics, welfare and the 3Rs (level 2); 11) Design of procedures and projects (level 2), (i) Legal issues; 24) Designated Veterinarian, (i) Legislation, (ii) Ethics, welfare and the 3Rs, (v) The principles of veterinary communications; (25) Project evaluator and (50) Local environment as well as Inspectors module.

**Syllabus:**

Topic 1: Introduction to LAS: Associations/organizations, professional networks

Topic 2: Legislation, including responsibilities of the LA veterinarian

Topic 3: Bioethics

Society perception of animal experimentation

Introduction to bioethics

Ethical frameworks

Bioethics applied to lab animals, including the 3 Rs

Topic 4: Concepts in laboratory animal welfare

Welfare and stress

Concept of animal welfare

Sentience

How can animal welfare be studied?

Overview of main welfare problems in lab animals

Physiology of stress

Main stressors in lab animals, including stress caused by transport

Positive re-enforcement, habituation/conditioning, preference tests and socialization of animals

Assessment of severity and humane end points

Pain and analgesia

Environmental enrichment and behavioural indicators of welfare

Re-use and the legal, practical and ethical issues on re-homing after use of animals in research

Retrospective evaluation

Topic 5: Assessment of protocols – general principles

Ethical committees

Cost-benefit analysis

Topic 6: Assessment of procedures – case studies

Rodents and rabbits

Carnivore

Pigs, ruminants and birds

Fish

NHP

Wild Animals

Topic 7: Inspection

**Module coordinador:**
Prof. Xavier Manteca Vilanova
Departamento de Ciencia Animal y de los Alimentos. Facultad de Veterinaria (UAB, Barcelona)

**Teaching Staff** (as per the last edition, March 2022):
David Anderson, Home Office, UK/ European Commission

Ignacio Álvarez Gómez de Segura, Veterinary School, University Complutense, Madrid, Spain

Marta Amat, Veterinary School of Barcelona, Spain

Aurora Bronstad, University of Bergen, Norway

Ana Criado, Aptuit, Verona, Italy

Anne-Dominique Degryse, ECLAM, France

Kate Garrod, Compliance Consultant, Red Kite Vets, UK

Jan Langermans, Biomedical Primate Research Center (BPRC). The Netherlands

Xavier Manteca, Veterinary School of Barcelona, Spain

Juan Ramos, Universitat Autònoma de Barcelona, Spain

Peter Sandoe, University of Copenhagen, Denmark

Hugh Simmons, Animal and Plant Health Agency (APHA), UK

Oriol Tallo Parra, Veterinary School of Barcelona, Spain

Patri Vergara, Veterinary School of Barcelona, Spain

Hanno Würbel, University of Bern, Switzerland

**Duration of Course:** 9 ECTS (44 attendance hours)

**Course dates:** 15 February - 30 April, 2025

**Attendance dates at UAB:** 1 week to be determined.

**Module:**

**Management of Animal Facilities. Breeding and Animal Care Programs.**

This module provides the knowledge and skills required for the effective management of animal facilities for experimental animals by designated veterinarians and other onsite personnel responsible for the welfare and care of laboratory animals.

The content of this module also complies with the training requirements set out in European Directive EU63/2010, developed by the Expert Working Group, supported by European Commission, in respect of the following modules: 3.1) Basic and appropriate biology (theory); 4) Animal care, health and management. 11) Design of procedures and projects (level 2) (iv) responsibilities ,23) Advanced animal husbandry, care and enrichment practices; 24) Designated veterinarian, (iii) Animal health, care and management.

**Syllabus:**

Topic 1. Strategic planning, impact of institutional policy and guidelines internal relations, relations with other animal centres and with society.

Topic 2. Animal facility design, construction and equipment: basic concepts; conforming to legislation and "good practices"; specialized functions of animals, projects, personnel and equipment.

Topic 3. Management of materials in animal facilities: hygiene and disinfection

Topic 4. Rodent facility: Equipment; management programmes, acclimatisation; environmental enrichment; security (access); automatic processes; analysis of specific needs and resolution of problems; equipment currently available; maintenance of

Topic 5. Rodent specialised facilities.

5.1 Digital cages.

5.2 Gnotobiotic models

Topic 6. Hamster and Guinea pig facility: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 7. Facility for rabbits: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 8. Non-Human Primates facility: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 9. Carnivore facility: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 10. Facility for swine: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 11. Ruminant facility: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 12. Facility for avian species: Equipment; management programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, bedding, waste disposal); equipment currently available; daily routines; animal identification.

Topic 13. Facility for aquatic species: Equipment; management programmes; breeding programmes; environmental enrichment; analysis of specific needs and resolution of problems (feeding, water quality); equipment currently available; daily routines; animal identification.

Topic 14. Health and Safety in animal facilities; legislation; health and safety procedures; evaluation and prevention of risks; protection of personnel; hygiene of the facility, control and disposal of waste.

Topic 15. Disaster planning

Topic 16. Human Resources.

16.1 Management of personnel. Categories of personnel and career programs; recruitment of personnel; incentives and evaluation of personnel.

16.2 Education and training in a research institution. Maintenance of personnel E&T portfolio.

Topic 17. Lean management: budgeting; purchasing policy; animal costing; obtaining animals- breeding or obtaining animals externally; effective financial management.

Topic 18. Breeding and animal stock management. Software available. Daily routines.

Topic 19. Handling, storing, traceability of data and samples.

Topic 20. Quality in animal care and use programmes.

Topic 21. Animal Transport. European Legislation.

PRACTICALS

Practical 1. Practical operations in IVCs

Practical 2. Digital cases. Data collection and analysis

Practical 3. Practical with equipment for individual protection

Practical 4. Visit to Animal Facility

**Programme Advisor:**

Javier Guillen, AAALAC International, Spain

**Teaching Staff** **(**as per the last edition, October 2022)**:**

Xavier Abad, CReSA-IRTA, Barcelona.

Isabel Blanco, CNIO, Madrid, Spain

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David Solanes Foz. Zoetis. Olot.Spain

Thomas Steckler, Janssen Research & Development, Belgium

Christian Urdiales Garcia, Techniplast S.p.A, Italy

Joana Visa, Visa Coaching Institute, Spain

Sarah Wolfensohn. School of Veterinary Medicine, University of Surrey, UK

**Duration of course:** 12 ECTS (80 attendance hours)

**Course dates:** 19 September -19 December, 2025

**Attendance dates at UAB:** one and a half weeks to be determined.

**Module:**

**Genetics of Laboratory Animals and Experimental Models**

This module provides the practical and theoretical knowledge and skills regarding genetics and experimental models required by designated veterinarians and other onsite personnel responsible for the welfare and care of laboratory animals in the conduct of experimental procedures.

The module consists of two parts. The experimental models’ component provides extensive knowledge of different experimental animal models in different research areas. The genetic component explores models of genetically modified animals, their production, maintenance and the implications for their use.

The content of this module also complies with the training requirements set out in European Directive EU63/2010, developed by the Expert Working Group, supported by the European Commission, in respect of the following modules: 3.2) Basic and appropriate biology (practical); 7) Minimally invasive procedures without anaesthesia (theory), 8) Minimally invasive techniques without anaesthesia (skills).

**Syllabus:**

Topic 1.Genetics, genetic control and genetically modified rodents.

1.1. Basic Concepts of the genetics of mammals.

1.2. Genetically defined lines of mice.

1.3 Genetic modification: transgenesis, targeted mutagenesis and genome editing. Tools and applications.

1.4. Genetically defined lines of rats and other rodents used in animal experiments. Spontaneous mutations. International standardized nomenclature of lines and mutations in rodents

1.5. Molecular Biology tools used in genetics of rodents

1.6. Genetic control and genetic contamination. Practical cases.

1.7. Standard nomenclature inbred, outbred congenic and mutant mice and rats

1.8. Standard nomenclature of GM mice

1.9. Rodent embryo manipulation. Collection, culture and transfer. Comparing mice and rats.

1.10. Genetic altered models in fish.

1.11 Management of colonies. Breeding strategies to keep consanguinity and avoid consanguinity. Breeding schemes in genetically altered animals

1.12 Influence of genetic background on phenotype, examples in neurobiology

1.13 Epigenetics, concept and impact on genotype and phenotype

1.14 Rederivation

1.15 The 3Rs in GA models. How to use international resources

1.16 Production and Management of GA colonies for Phenotyping Platforms

1.17 Rodent basic phenotyping

1.18 Phenotypes by image

 1.19 Phenotypes by behaviour

1.20 Reproductive biotechnology. Cryopreservation. FIV

1.21 From founder to line: Establishing a Genetically Altered (GA) line for research purposes

1.22 Establishing a new genetically altered line according to Directive 2010/63/EU. Practical issues.

1.23 Rat husbandry differences in relation to mice

1.24. Practical sessions:

1.24.1. Genotyping procedures

1.24.2. Abdominal and scrotal vasectomy

1.24.3. Obtaining sperm and FIV

1.24.4. Cryopreservation 2 cell and morulae

1.24.5. Collection and Transfer of mouse1-cell embryos

1.24.6. Phenotyping behaviour

Topic 2. Selection of animal model according to biological characteristics

2.1. Biology, management and care guidelines in cephalopods

2.2 Nutrition and its influence on research

2.3 Practical sessions on management, sexing, marking, administration and taking samples in the main species of experimental animals:

2.3.1 Mice and rat

2.3.2 Rabbits

2.3.3 Pigs

2.3.4 Ruminants

2.3.5 Birds

2.3.6 Fish

2.3.7 Amphibians

2.3.8 Cephalopods

2.3.9 Dogs (demonstration)

2.3.10 Nonhuman primates (demonstration)

Topic 3 Selection of experimental model according to type of research.

3.1. Introduction to animal models

3.2 Experimental models in neurodegenerative and rare diseases

3.3 Experimental models in cardiovascular diseases

3.4. Experimental models in oncology.

3.5 Experimental models in diabetes and metabolic diseases.

3.6 Experimental models in Reproduction

3.7 Experimental models in foetal medicine

3.8 Experimental models in immunology.

3.9 Experimental models in infectious diseases.

**Module Coordinators:**

Belen Pintado, Centre Nacional of Biotechnology, Madrid, Spain

Sergio Berdún Marín, Research Parc of Barcelona (PCB) and Veterinary School, Barcelona, Spain

**Teaching Staff** **(**as per the last edition, March 2023)**:**

Lucía Acal, HIPRA, Amer, Spain

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Eloi Barcelona, HIPRA, Amer, Spain

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Verónica Domínguez Plaza, CNB-CSIC, Madrid Spain

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Graziano Fiorito, Stazione Zoologica Anton Dohrn, Napoli, Italy

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Teresa Paramio, Departament de Ciència Animal, Universitat Autònoma de Barcelona Joan Perez, HIPRA, Amer, Spain

Belén Pintado. Centro Nacional de Biotecnología, CNB-CSIC, Madrid. Jordi Polo, Anapath Research, Spain

**Duration of course:** 15 ECTS, 88 attendance hours

**Course dates:** 15 February -15 May 2026

**Attendance dates at UAB:** two weeks to be determined.

**Module:**

**Microbiology, Diseases and Health Monitoring**

This module provides the practical and theoretical knowledge and skills regarding microbiology, diseases and health monitoring required by designated veterinarians and other onsite personnel responsible for the welfare and care of laboratory animals in the conduct of experimental procedures.

The module looks at the importance of health monitoring and the range of diseases that can affect laboratory animals and how diseases may affect experiment results.

This module also provides practical training in relation to animal biology, euthanasia and minimally invasive methods.

**Syllabus:**
Topic 1. Introduction to the pathology and infectious diseases of laboratory animals

Topic 2. Definition, importance and microbiological characteristics of laboratory animals. Microbiota and its influence in animal models.

Topic 3. Diseases in rodents and rabbits.

Basic concepts.

Diseases in mice.

Diseases in rats.

Diseases in guinea pigs.

Diseases in rabbits.

Topic 4. Health monitoring in rodents and rabbits

Importance of infectious diseases in animal experimentation

Health monitoring following FELASA recommendations

Evaluation of the risk of spread of infectious agents in animal housing facility

Maintenance and control of a high quality sanitary status

Practical case study

Topic 5. Clinical biochemistry and hematology studies

Basic concepts and interpretation of results

Topic 6. Clinical sings in experimental animals

Topic 7. Microbiology and diseases in carnivores

Diseases in dogs, cats and ferrets

Health monitoring: FELASA recommendations.

Prevention of diseases, clinical examination and taking samples.

Topic 8. Microbiology and diseases in fish

Introduction

Microbiology and diseases in fish

Health monitoring and biosecurity in laboratory fish: FELASA recommendations

Topic 9. Microbiology and diseases in ruminants and pigs

Microbiology and diseases

Health monitoring: FELASA recommendations

Topic 10. Microbiology and diseases in primates

Health monitoring: FELASA recommendations.

Disease Prevention, clinical exploration, sampling.

Primate diseases

Topic 11. Microbiology and diseases of poultries

Health monitoring

Disease Prevention, clinical exploration, sampling.

Primate diseases

Topic 12. Experimental pathology

Practical sessions: Necropsy and tissue sampling in rodents and pig.

**Module coordinator:**  M Neus Prats Costa. IRB Barcelona – Institute for Research in Biomedicine, Barcelona, Spain

**Teaching Staff**

Mariona Aulí, Almirall, Spain

Marion Berard, Institute Pasteur, France

Laurence Bonnet, Marshall BioResources, France

Stephanie Buchheister , Hannover Medical School, Germany

Mariano Domingo, Departament of Animal Health. Universitat Autònoma de Barcelona, Spain

Ricardo Feinstein, The National Veterinary Institute, Sweden

Patrick Hardy, FELASA Working Group on Farm Animals, France

Jan Langermans, Biomedical Primate Research Center (BPRC), The Netherlands.

Ricard March, Laboratorios Hipra S.A., Spain

Jean-Philippe Mocho, Joint Production System Ltd., UK

Francesc Padrós, Departament of Animal and Vegetal Biology and Ecology. UAB, Spain

Neus Prats, Institute for Research in Biomedicine IRB-Barcelona, Spain

Irene Ruano, Institute for Research in Biomedicine IRB-Barcelona, Spain

Yolanda Saavedra, University of Surrey, UK

**Duration of course:** 6 ECTS (44 attendance hours)

**Course dates:** 1 October - 15 December 2023

**Attendance dates at UAB:** 20-24 November 2023

**Module:**

**Anaesthesia, Analgesia and Euthanasia of Laboratory Animals**

This module covers anaesthesia, analgesia and euthanasia of the main species of laboratory animals and their impact on animal welfare and experimental results. The course also focuses on the knowledge required by designated veterinarians and other onsite personnel responsible for the welfare and care of laboratory animals in the conduct of experimental procedures.

The content of this module complies with the training requirements set out in the European Directive EU63/2010 developed by the European Commission in respect of the following modules: 6.1 Humane methods of killing (theory); 6.2 Humane methods of killing (skills); 20 Anaesthesia for minor procedures; 21 Advanced anaesthesia for surgical or prolonged procedures; 24 Designated Veterinarian, (iv) Anaesthesia, analgesia, surgery.

**Syllabus:**

Anaesthesia

Topic 1. Basic principles (review)\*

1.1 Peri-operative management. Fluidotherapy

1.2 Pharmacology of anaesthetics. Drug selection

1.3 Anaesthetic equipment and breathing circuits

1.4 Monitoring

1.5 Drug and animal data management

Topic 2. Anaesthesia in most common species

2.1 Anaesthesia in rodents

2.2. Anaesthesia in rabbits

2.3. Anaesthesia in pigs

2.4. Anaesthesia in ruminants

2.5 Anaesthesia in fish, birds and reptiles

2.6. Anaesthesia in primates

2.7. Anaesthesia in carnivores (review)\*

Topic 3. Mechanical ventilation

3.1 Basic mechanical ventilation

3.2 Advanced mechanical ventilation

Topic 4. Local and regional anaesthesia

4.1 Local and regional anaesthesia

4.2 Electro & ultrasound guided regional anaesthesia

Topic 5. Monitoring

5.1 Monitoring in rodents

5.2 Advanced Monitoring

Topic 6. Impact of anaesthesia in research results

Analgesia

Topic 7. Recognising and Alleviating Animal Pain

Topic 8. Management of Pain in laboratory animals

Topic 9. Analgesia in non-mammalian species

Euthanasia

Topic 10. Update on euthanasia of laboratory animals

Practical Sessions

Anaesthesia in rodents and rabbits

Anaesthesia in sheep, including advanced ventilation

Anaesthesia in pig, including electrical-guided and ultrasound-guided techniques

Simulation of anaesthesia, cardiovascular complications

**Module Coordinator:**

Ignacio Álvarez Gómez de Segura. Facultad de Veterinaria, Universidad Complutense, Madrid.

**Teaching Staff (**as per the last edition, March 2021)**:**

Klas Abelson. Department of Experimental Medicine, University of Copenhagen, Denmark

Adrià Aguilar. Faculty of Veterinary Medicine, Universitat Autonoma de Barcelona, Spain

Ignacio Álvarez. Faculty of Veterinary Medicine, Universidad Complutense de Madrid, Spain

Mario Arenillas. Getafe Hospital, Madrid, Spain

Javier Benito. Faculty of Veterinary Medicine, University of Montreal, Canada

Susana Canfran. Faculty of Veterinary Medicine, Universidad Complutense de Madrid, Spain

Cristina Costa. Faculty of Veterinary Medicine, Universitat Autonoma de Barcelona, Spain

Paul Flecknell. Comparative Biology Centre. University of Newcastle, UK

Patricia Hedenqvist. University of Agricultural Sciences. Sweden

Francisco Laredo. Faculty of Veterinary Medicine, Universidad de Murcia, Spain

Carlota Largo. Hospital la Paz. Madrid, Spain

Xavier Moll. Faculty of Veterinary Medicine, Universitat Autonoma de Barcelona, Spain

**Duration of course:** 6 ECTS (44 attendance hours).

**Course dates:** 1 March - 15 May, 2024

**Attendance dates at UAB**: 11-15 March, 2024

**Module:**

**Reduction and Refinement in Experimental Design and Methodology. Research and Development of Drugs**

This module provides the practical and theoretical knowledge and skills regarding experimental design and methodology required by designated veterinarians and other onsite personnel responsible for the welfare and care of laboratory animals in the conduct of experimental procedures.

This course covers the most important aspects in relation to the reduction in the number of animals used in research, experimental design, literature searches and presentation of results. It also deals with refinement through the study of the main non-invasive and minimally invasive techniques and refinement in surgical techniques. The module also provides an overview of the strategy and processes required in research and drug development.

The content of this course also complies with the training requirements set out in European Directive EU63/2010 developed by the European Commission in respect of the following modules: 10. Design of procedures and projects (level 1), 11. Design of procedures and projects (level 2), 22. Principles of surgery and 51. Information provision and retrieval.

Block I: Reduction

Topic 1. Experimental design

Topic 2. Systematic Reviews

Topic 3. Internal validity and bias analysis

Block II: Refinement

Topic 4. Asepsis and preparation for surgery. Aseptic surgery in rodents

Topic 5. Pre-operative assessment, perioperatory and postoperatory care.

Fluidotherapy and antibiotherapy

Topic 6. Microsurgery techniques: vascular surgery

Topic 7. Blood sampling techniques

Topic 8. Recording of physiological parameters by non-invasive and minimally invasive techniques

Topic 9. Imaging techniques

9.1. Bioluminiscence and fluorescence

9.2. Magnetic Resonance Imagining (MRI)

9.3. Positron Emission Tomography (PET)

9.4 Echography

9.5 Micro Computed Tomography

Block III: Experiments under legal requirements

Topic 10. Drug development and Drug Efficacy

Topic 11.Development of veterinary drugs

Topic 12. Drug development: Safety and toxicology

Topic 13. Biologicals: immunogenicity and immunotoxicity

Topic 14: Translational research

Topic 15: Quality standards: Good Laboratory Practice and consistency in conducting scientific procedures and correct handling, storing, recording, and ensuring traceability of samples

P R A C T I C A L S E S S I O N S

Practical session 1.1: Imaging techniques in rodents: Positron Emission Tomography,

Ecography. MicroCT, Bioluminiscence and Biofluorescence (IVIS).

Practical session 1.2: Imaging techniques: Magnetic Resonance Imaging (MRI).

Practical session 2.1: Principles of surgery in refinement. Asepsis. Sutures and knots. Aseptic surgery in rats. Minimally invasive devices

Practical session 2.2: Introduction to microsurgery.

Practical session 2.3: Cannulation, catheters.

Practical session 2.4: Laparoscopy and Ultrasound guided vascular cannulation

Practical session 2.5: Blood sampling techniques in rodents

Practical session 3.1: Telemetry

Practical session 3.2: Plethysmography.

**Module coordinador:** Joan Antoni Fernández Blanco. Parc de Recerca Biomèdica de Barcelona (PRBB) Barcelona and Faculty of Veterinary Medicine, Universitat Autònoma de Barcelona

**Teaching Staff** (as per the last edition, October 2021):

Julio Alvarez, Data Sciences International, Belgium

Elena Abellán Rubio, Microsurgery unit, Centro de Cirugía de Mínima Invasión Jesús Usón, Cáceres, Spain

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Carlos Baldellou Estrada, Universitat Autònoma de Barcelona, Barcelona, Spain

Manuel Berdoy, Oxford University, UK

Marco Boscaro, Data Sciences International, Italy

Juan Antonio Cámara, Helen Diller Family Comprehensive Cancer Center, University of California San Francisco, USA

Ana Paula Candiota, Centro de Investigación Biomédica en Red, Department of Molecular biology and Biochemistry, Universitat Autònoma de Barcelona, Spain

Jaime Donate Galiano, Preclinical Imaging Platform, Lab Animal Service, Vall d’Hebron Research Institute (VHIR), Barcelona, Spain

Marielle Esteves Coelho, Vall d’Hebron Research Institute (VHIR), Barcelona,

Spain

Laia Foradada, Peptomyc S.L. / Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain

Laura Fresno, Endolab, Hospital Clinic Veterinari, Barcelona, Spain

César Galo García Fontecha, Hospital Sant Joan de Deu, Barcelona, Spain

Marta Giral, Almirall, Barcelona, Spain

Francisca Gomez, Anapath Research, Barcelona, Spain

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Silvia Lope, Servei de Ressonància Magnètica Nuclear (Service of MRI),

Universitat Autònoma de Barcelona, Barcelona, Spain

Stephan Martin, Data Sciences International, Belgium

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Fernando de Mora, Department of Pharmacology, Universitat Autònoma de Barcelona, Barcelona, Spain

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Kate Read, Labcorp Drug Development, Huntingdon, UK

Daniel Ruiz, Trinity College, Dublin, Ireland

David Sabaté, Hipra Laboratories, Amer, Spain

Beatriz Salinas Rodríguez, Instituto de Investigación Sanitaria Gregorio Marañón,

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Laura Santos, Endolab, Hospital Clinic Veterinari, Barcelona, Spain

Anna Server Salvà, Vall d’Hebron Research Institute (VHIR), Barcelona, Spain

Marcello Tontodonati, GlaxoSmithKline, Hertforshire, UK

Francisco Javier Vela, Microsurgery unit, Centro de Cirugía de Mínima Invasión Jesús Usón, Cáceres, Spain

Kim Wever, Radboud University Medical Center, Radboud Institute for Health Sciences, Dept. of Anesthesiology, Nijmegen, The Netherlands

**Duration of course**: 12 ECTS (80 attendance hours)

**Course dates**: 1 October -15 December, 2024.

**Attendance dates at UAB**: 10 days to be determined

**Practicum**

This training, accounting for 15 ECTs, takes place in the student's own workplace and is supervised by Master course teachers. The training is supplemented with placements in other animal centers which satisfy the requirements for practical training for the Master course.

Each student must prepare a detailed written description of their workplace and their tasks performed. The student's Master tutor will establish a plan for monitoring and evaluating the student's work. Depending on the amount of time that the student is able to dedicate to specific tasks, the variety of species housed in their centre and the diversity of tasks, the Master coordinator may assign additional tasks to supplement the training.

To ensure that students have experience in all areas of the Master syllabus, students are required to complete a practical placement of approximately one month duration, in a different animal centre to the one they work in focusing on areas in which they need experience. This placement may be divided into two to facilitate compatibility with student’s job obligations.

**Project**

The Masters course project comprises two components:

 1. A report detailing the student's practical experience to date in relation to laboratory animal science including the experience obtained at work, from the Master's practicum as described above and from the practical sessions on the Masters course.

2. The second requirement of the project is to demonstrate accredited experience in laboratory animal research by presenting an original research paper as first author which must be published in a journal listed in the Science Citation Index (SCI)

The research project may be conducted before, during or after completion of the modular training and work experience placements and the research may be done in any European or non-European research centre.

Once completed, the research paper and practical experience report are submitted for assessment by a committee of the master’s teaching staff.

**Current fees for 2023-2026**

**Ordinary fee master complete to be paid in 3 academic years: 18,235 €**

**Personnel from public research centres: 16,440€**

**Awardees of scholarship for the master:** **15,500€**

**Members of SECAL, ESLAV and ECLAM Residents: 15,500 €**

**Price per module:**

**Module 1: 1,750 €**

**Module 2: 2,475 €**

**Module 3: 3,960 €**

**Module 4: 1,950 €**

**Module 5: 2,550 €**

**Module 6: 3,570 €**

**Fee per module may have a 15% discount to members of SECAL, ESLAV and ECLAM Residents.**

**Sponsors**

The course thanks **Serveis Integrats de l’Animal de Laboratori (SIAL)**

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**International Council for Laboratory Animal Science (ICLAS)**



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for their support with individual scholarships.